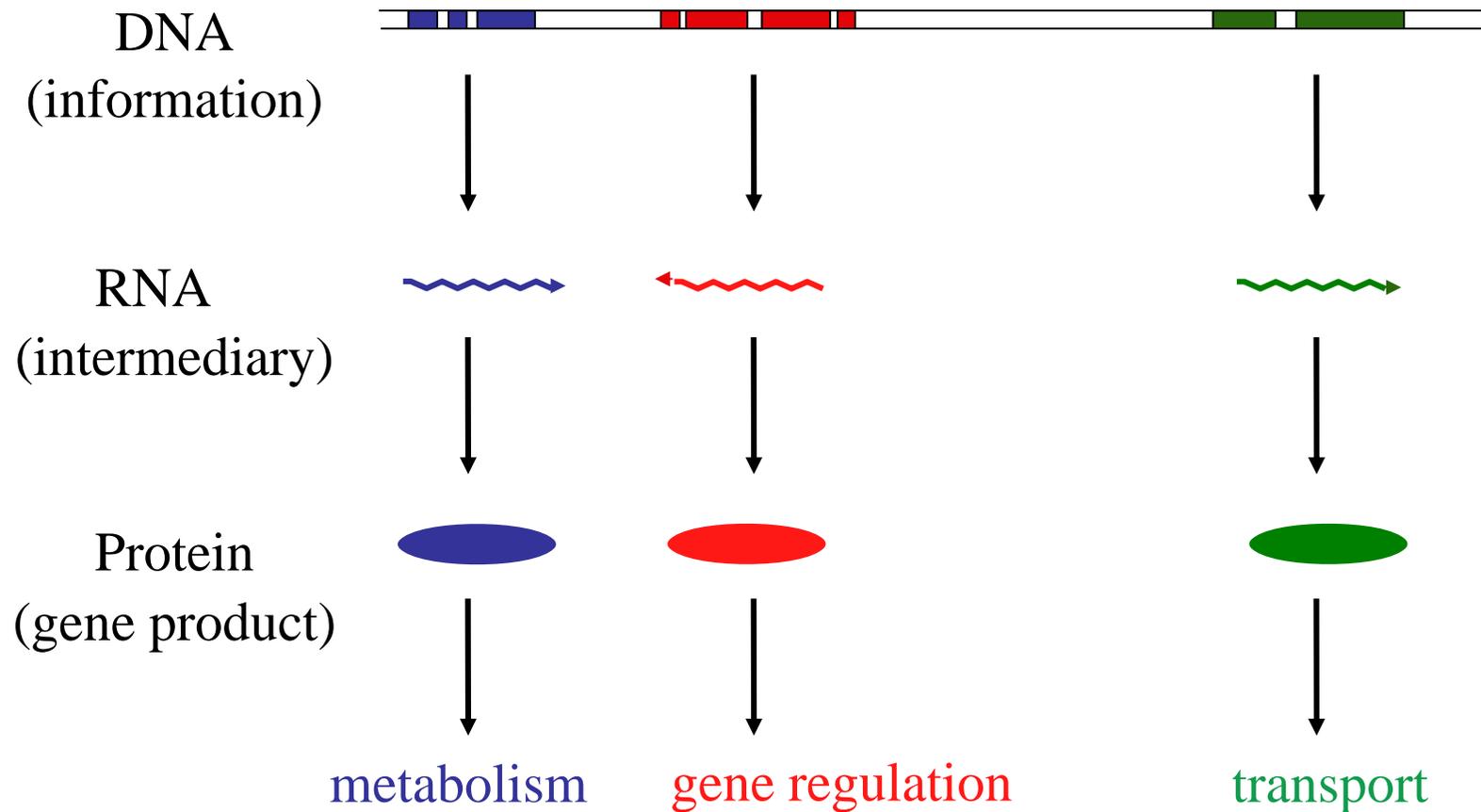
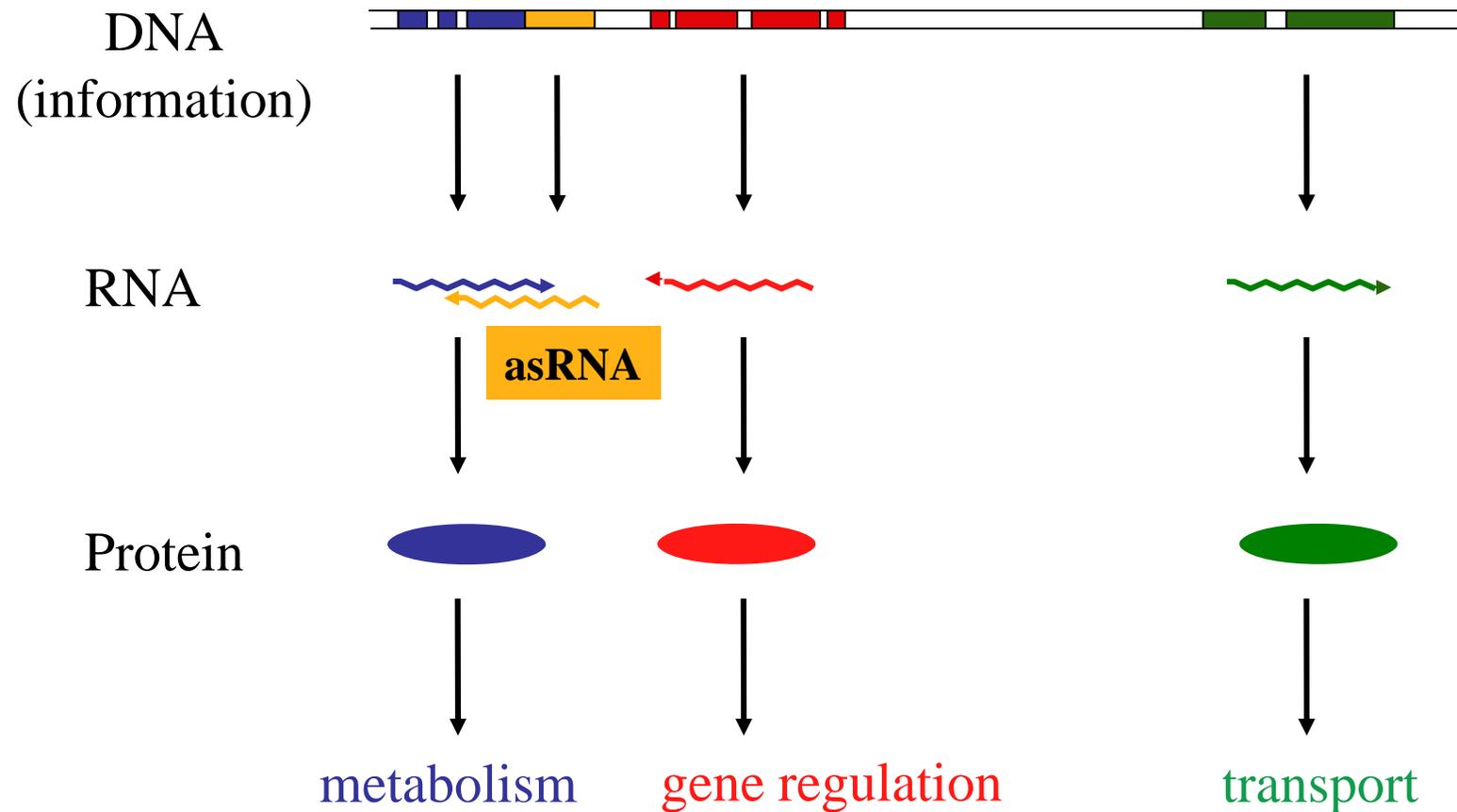


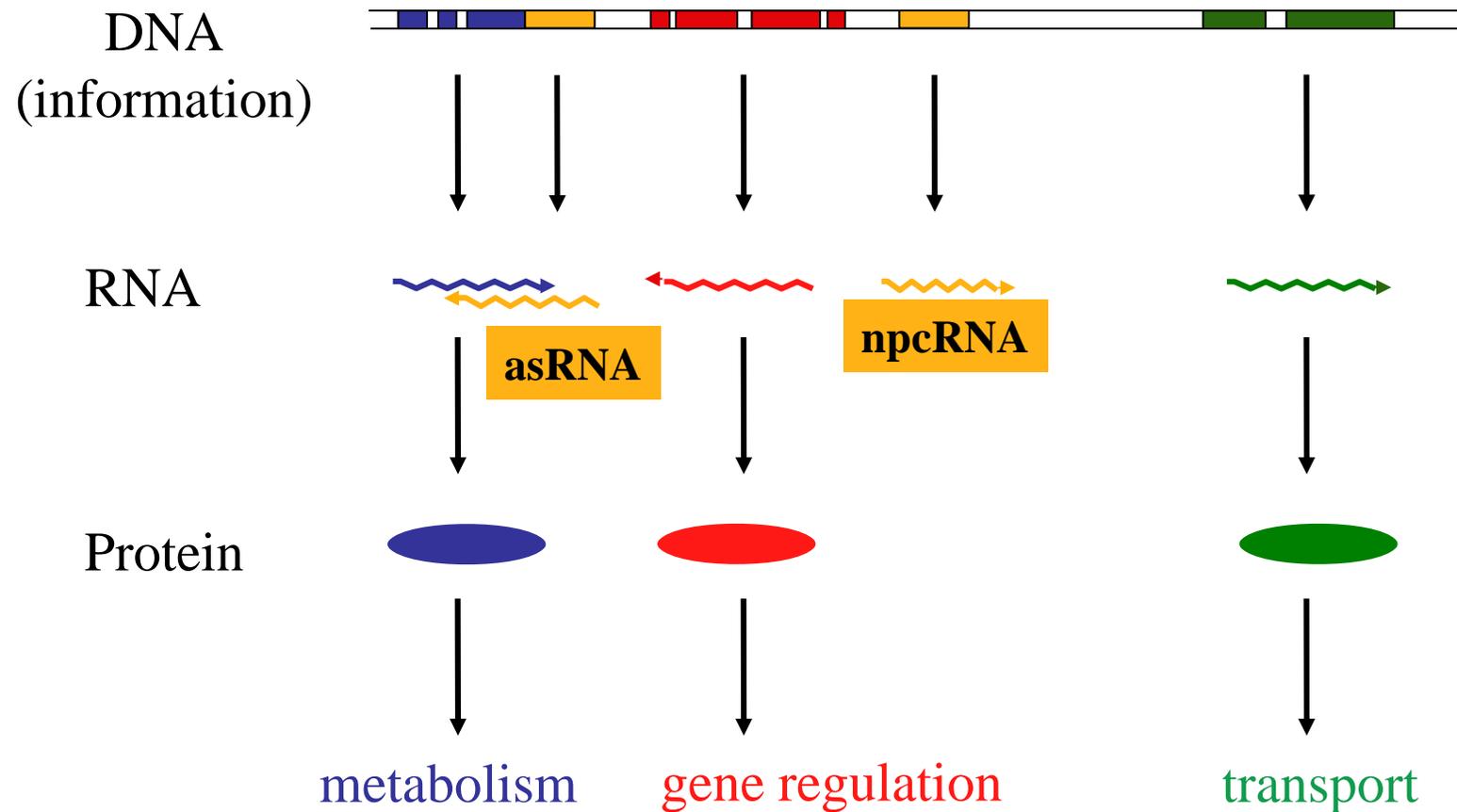
The genome contains more than protein coding genes



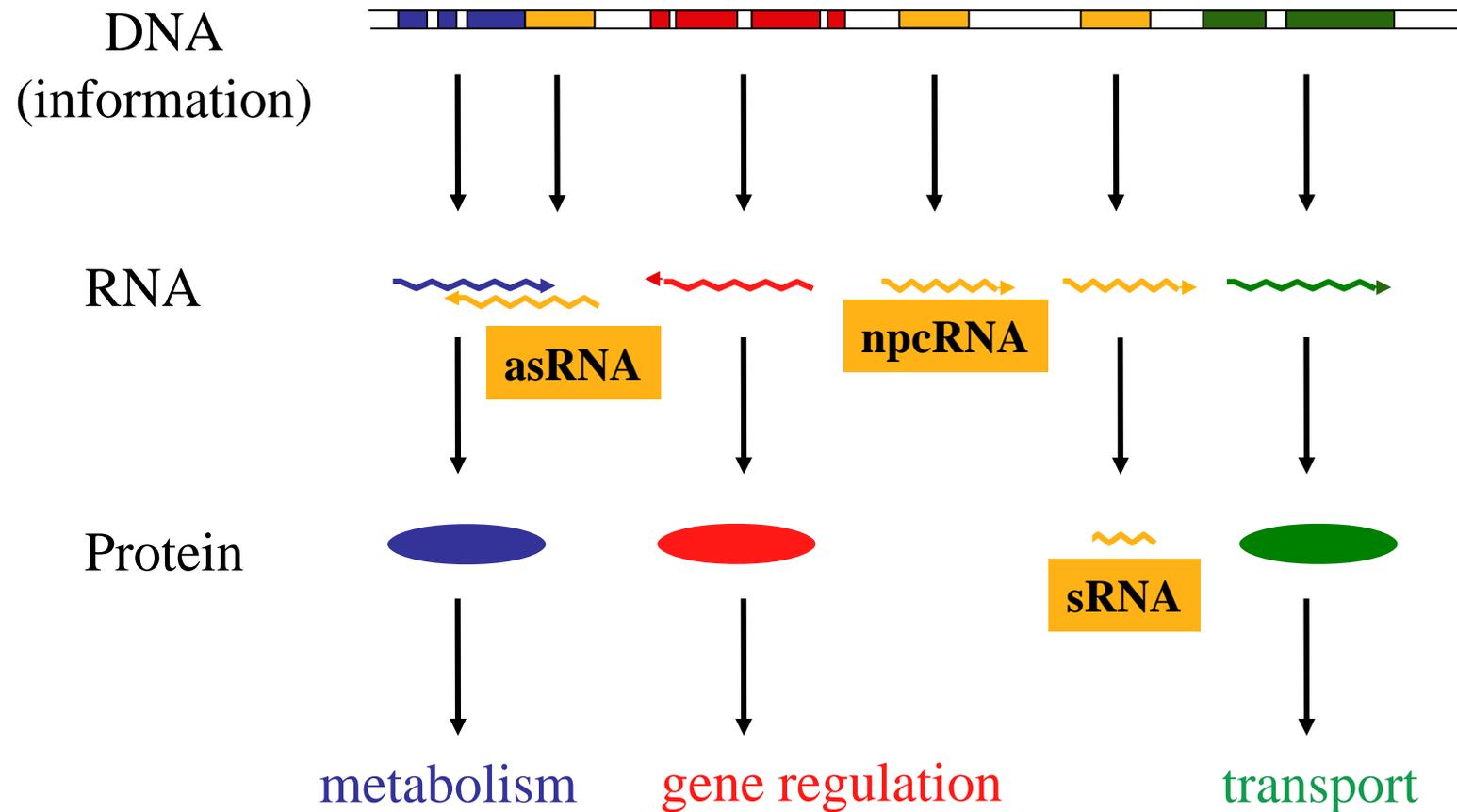
At least three types of **non-protein coding genes** exist



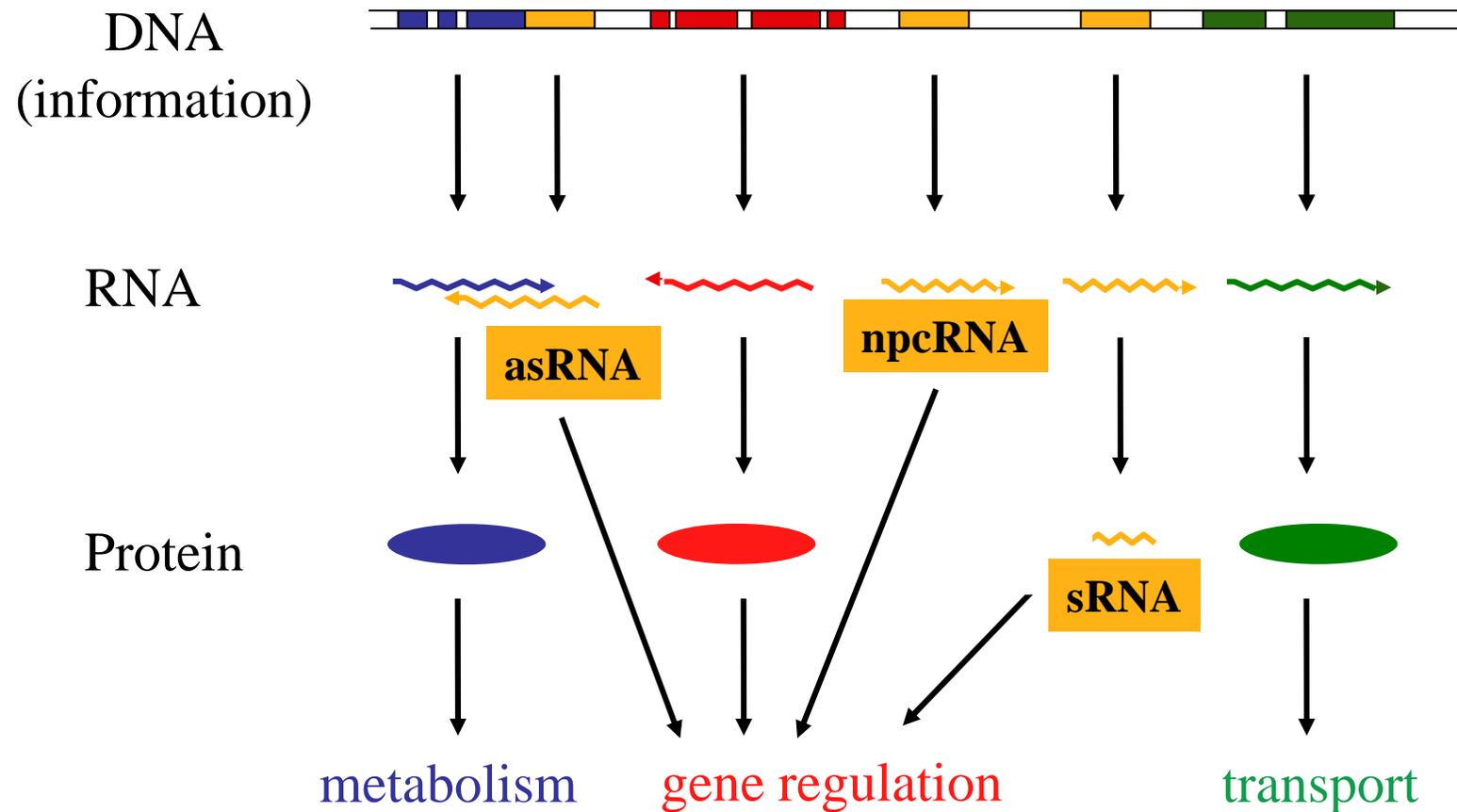
At least three types of **non-protein coding genes** exist



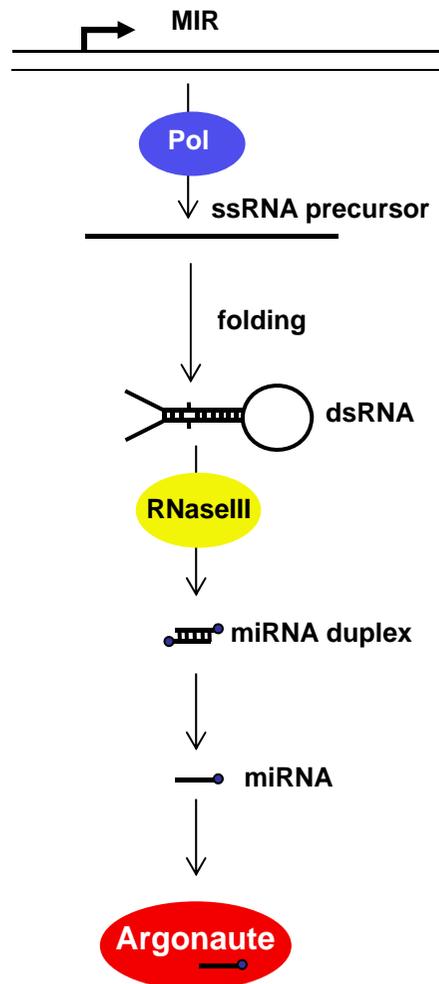
At least three types of **non-protein coding genes** exist



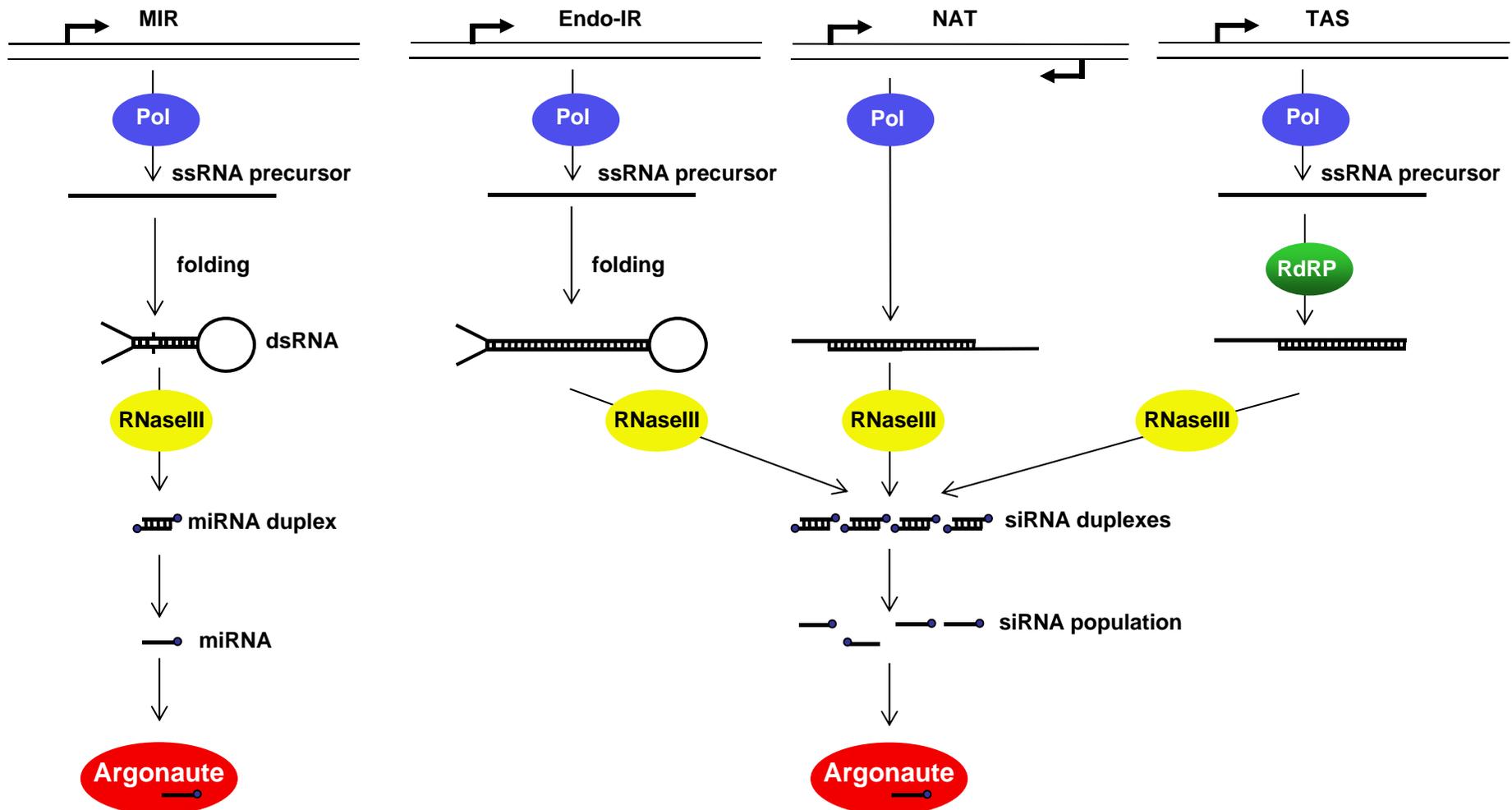
At least three types of **non-protein coding genes** exist



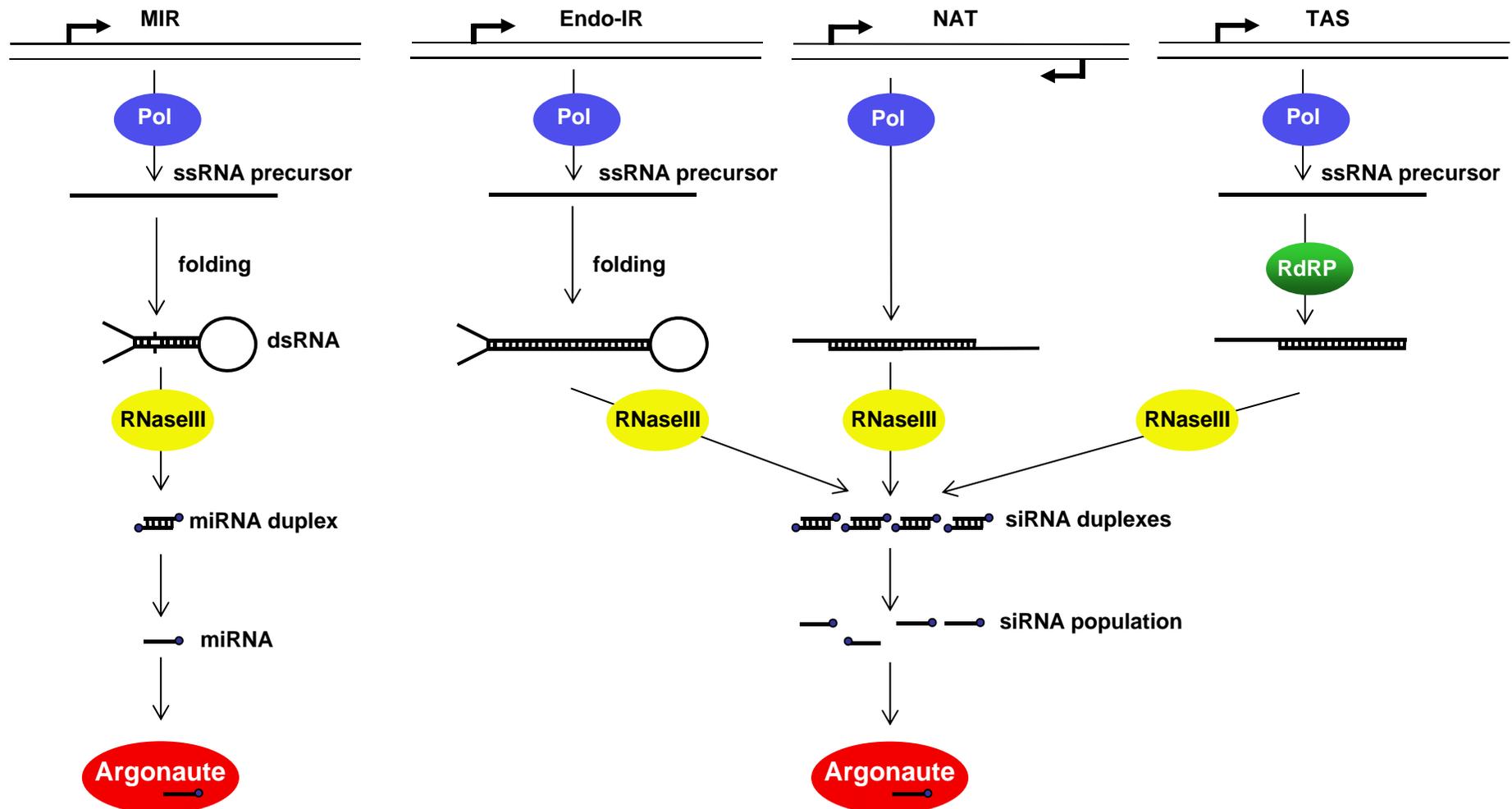
Plants contain miRNA and multiple types of siRNA



Plants contain miRNA and multiple types of siRNA



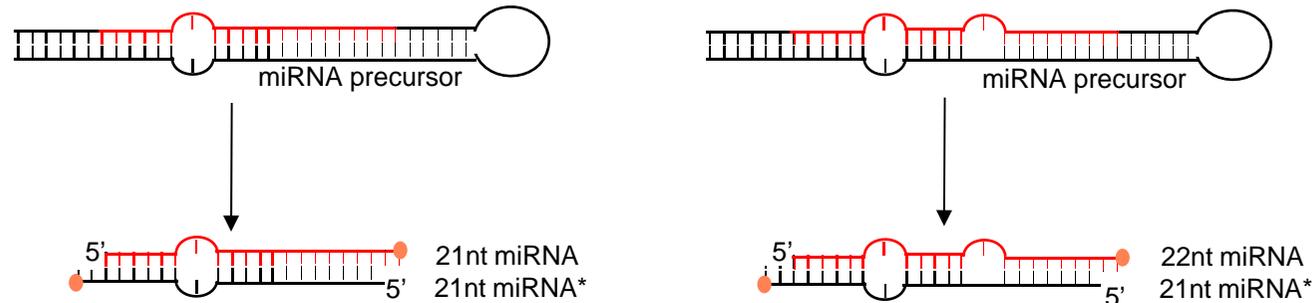
Plants contain miRNA and multiple types of siRNA



21-22-nt mi/siRNA guide PTGS while 24-nt siRNA guide TGS

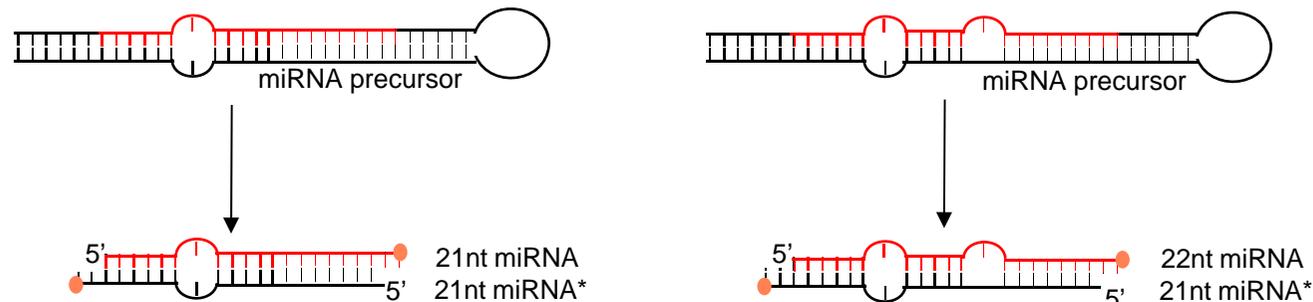
Different DCL produced small RNA of different sizes

DCL1 -> 21-nt miRNA (19-25-nt depending on the structure of the stem-loop)



Different DCL produced small RNA of different sizes

DCL1 -> 21-nt miRNA (19-25-nt depending on the structure of the stem-loop)

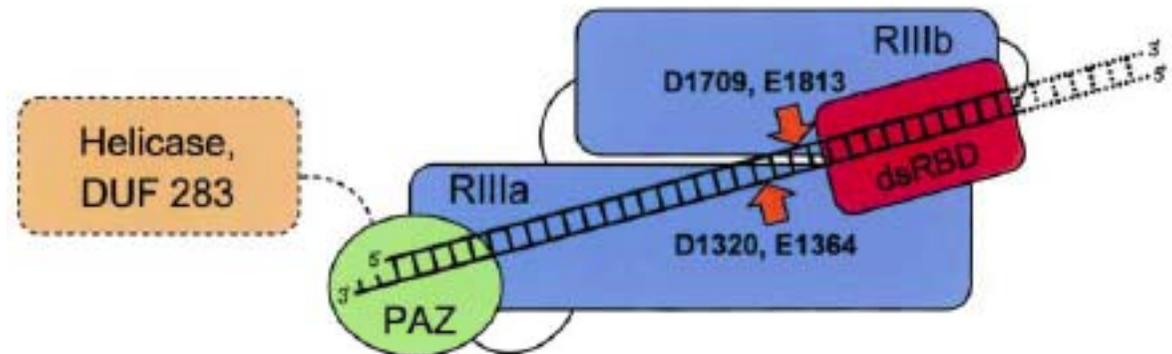


DCL2, DCL3 and DCL4 act as rulers

DCL4 -> 21-nt siRNA

DCL2 -> 22-nt siRNA

DCL3 -> 24-nt siRNA



DCL1 is essential for plant development



Wild-type



null *dcl1*



hypomorphic *dcl1*

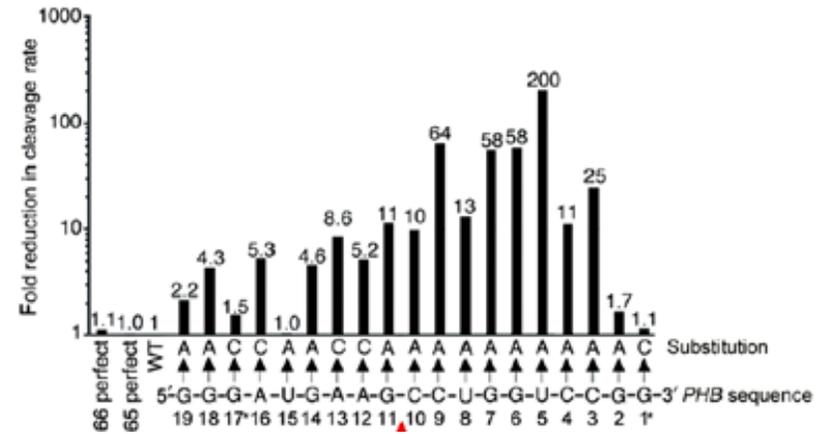
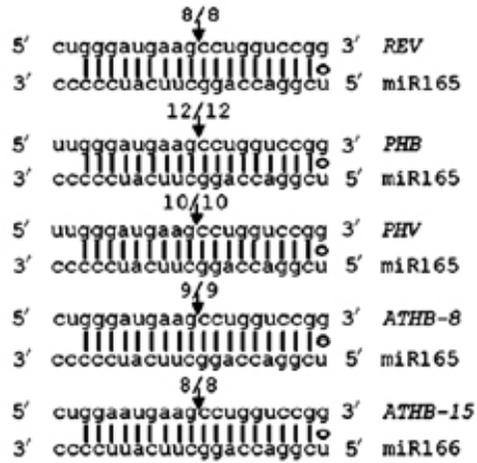
miRNAs guide translational repression

In animals, miRNA regulate thousands of targets by translational repression.

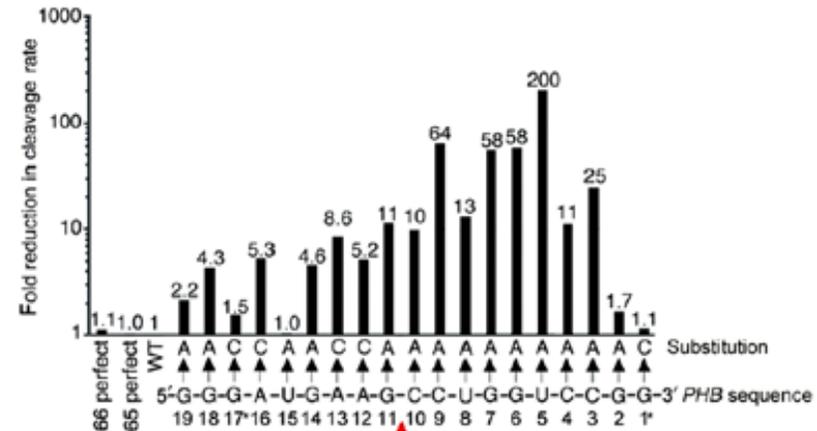
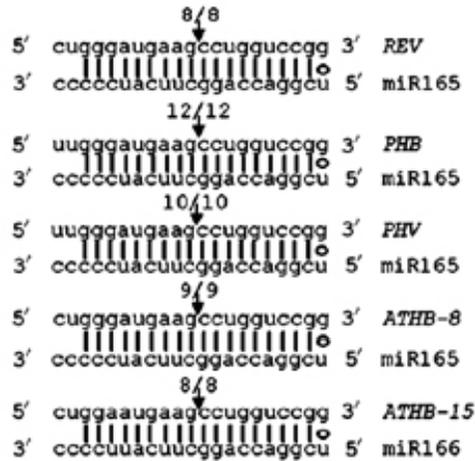
In plants, only 4 miRNA targets regulated by translational repression have been identified. However, no systematic proteomic analyses have been performed.

-> many more targets may exist in plants

miRNAs guide target RNA cleavage



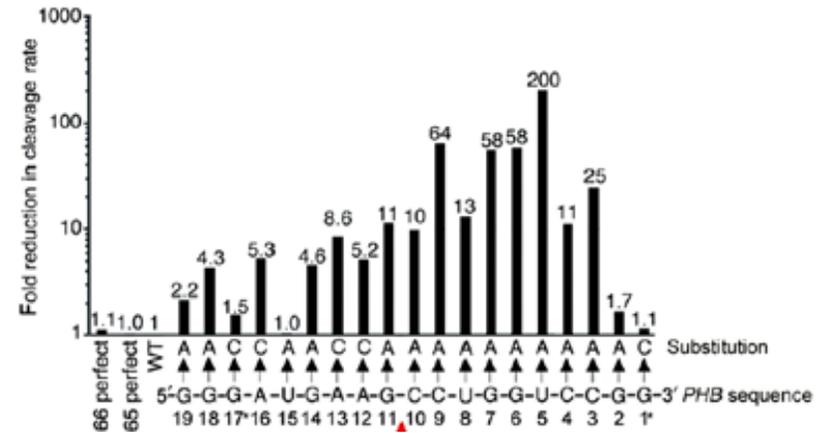
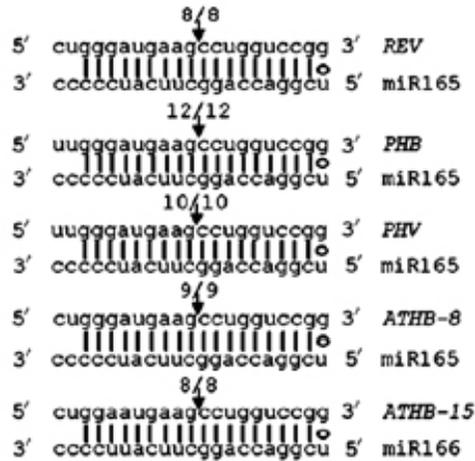
miRNAs guide target RNA cleavage



Algorithms allowing up to 3 mismatches confidently predicted hundreds of miRNA targets regulated by cleavage in plants

However, bioinformatics is insufficient to predict all miRNA targets

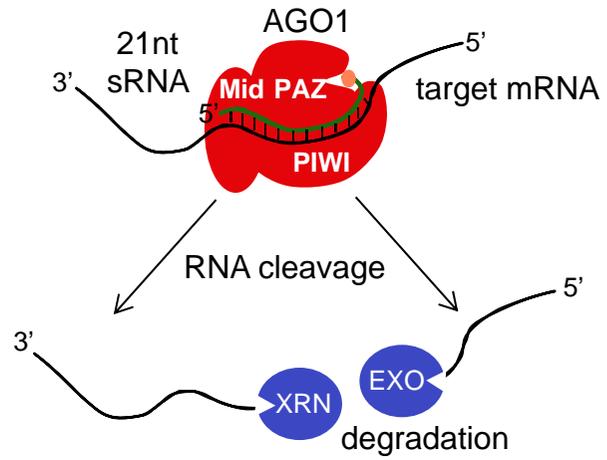
miRNAs guide target RNA cleavage



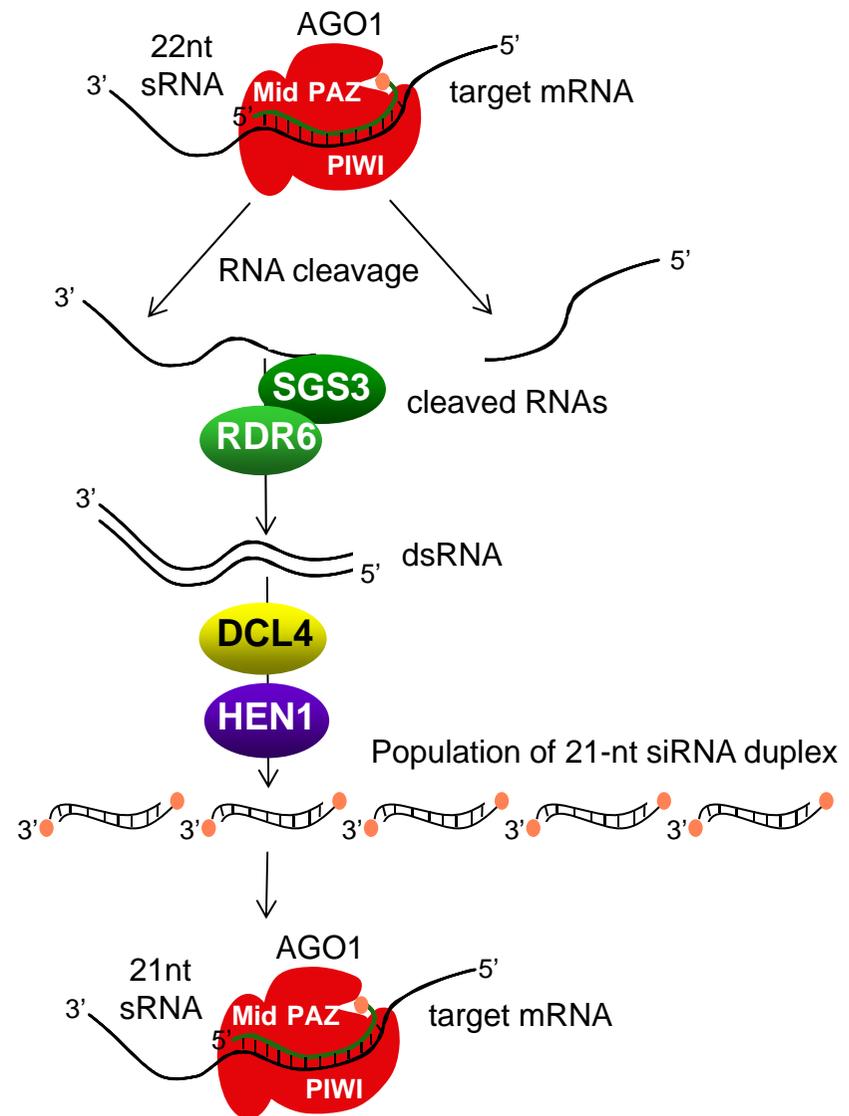
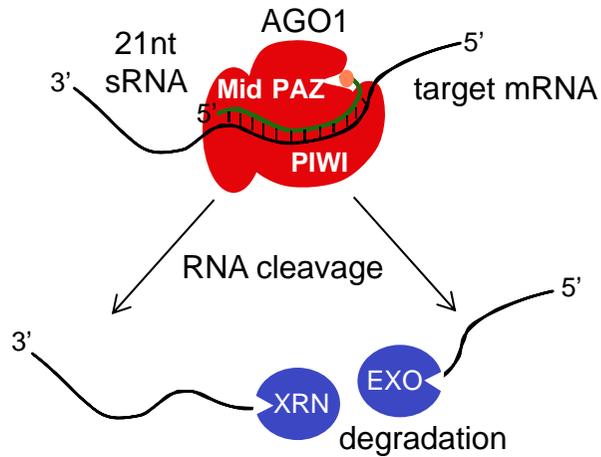
Genetics identified miRNA targets with up to 5 mismatches



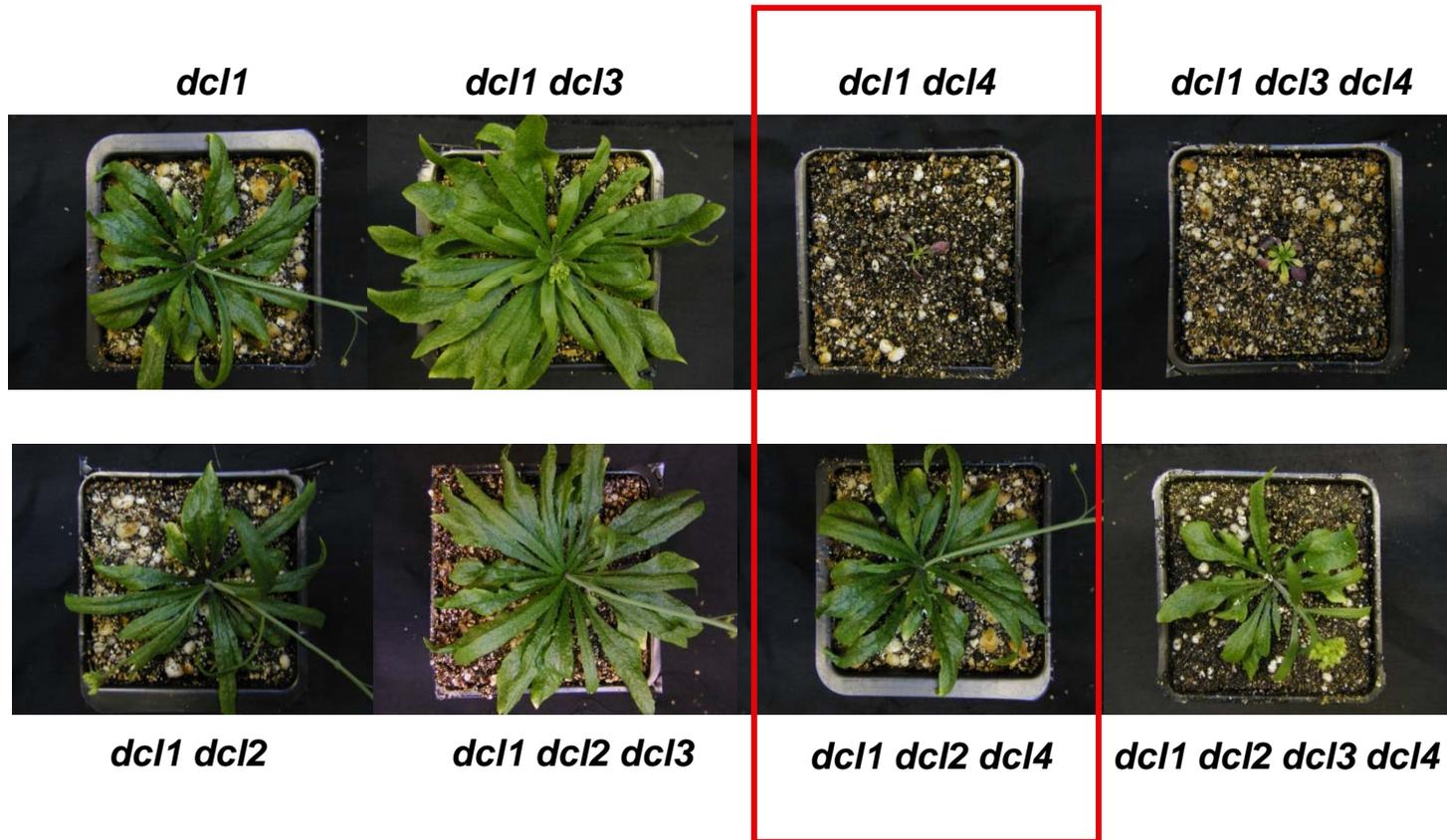
21-nt-guided cleavage products are degraded



21-nt-guided cleavage products are degraded while 22-nt-guided cleavage products produce secondary siRNA

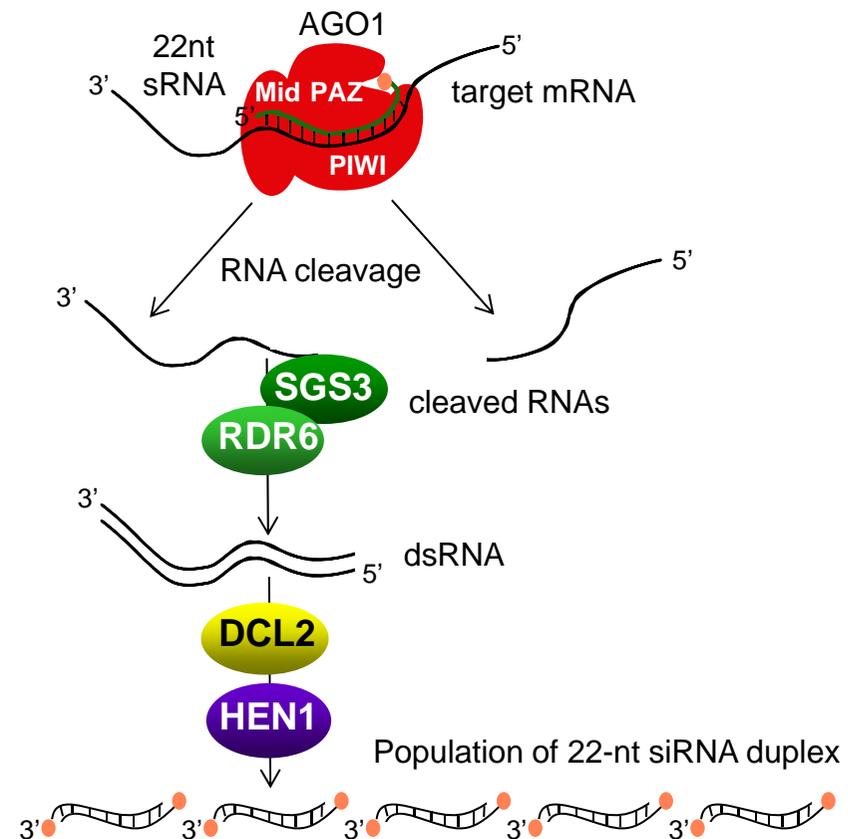
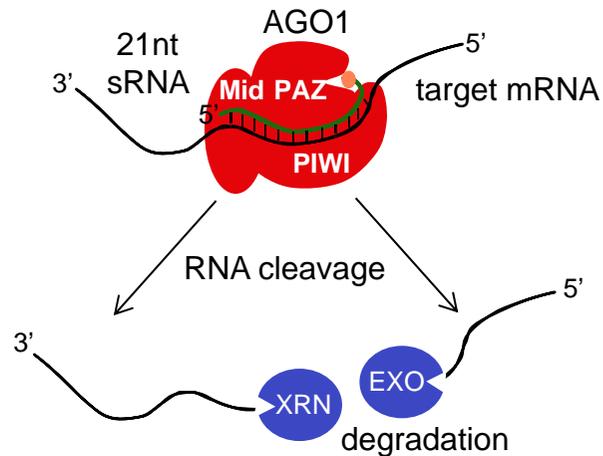


DCL2 : one nt size change makes a big difference



DCL2 imperfectly substitutes to DCL4 and produces 22-nt instead of 21-nt

21-nt-guided cleavage products are degraded while 22-nt-guided cleavage products produce secondary siRNA



In *dcl1 dcl4*, which lacks 21-nt siRNAs, 22-nt siRNAs made by DCL2 promote secondary 22-nt siRNAs, which promote tertiary 22-nt siRNAs, which promote...

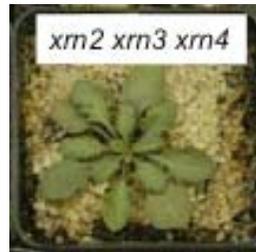
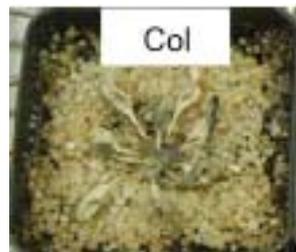
XRN and Exosome are essential for plant development



null *xrn3*

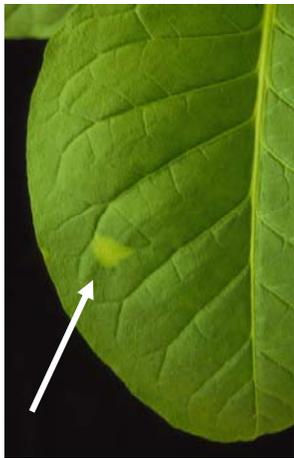


null *rrp4*



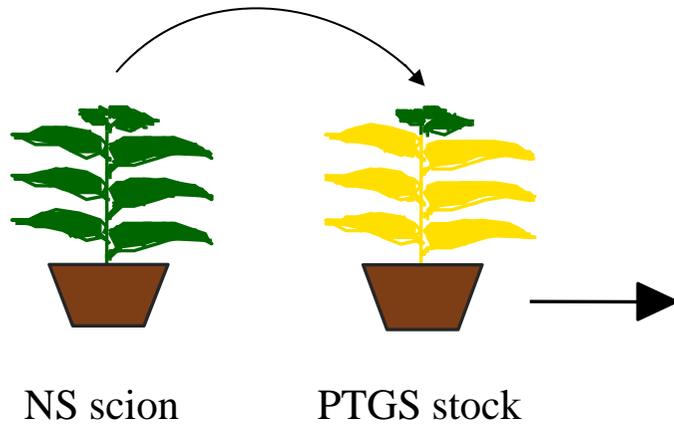
Partial impairment of the XRN pathway promotes drought stress resistance

siRNA of all sizes are mobile

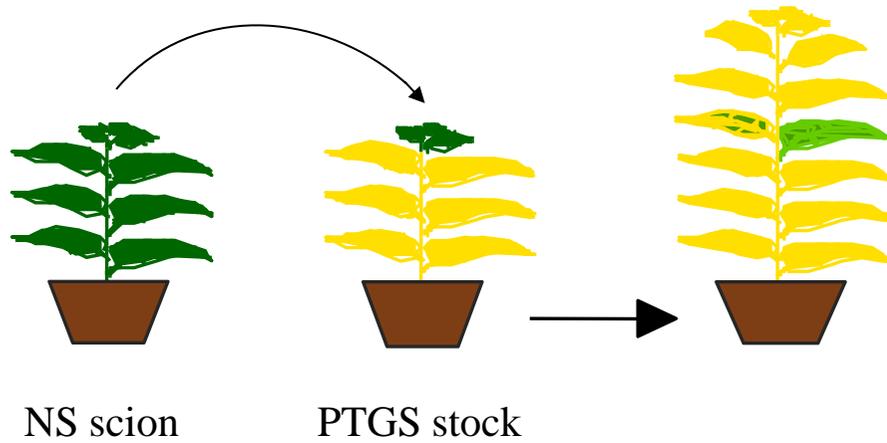


Progression of silencing

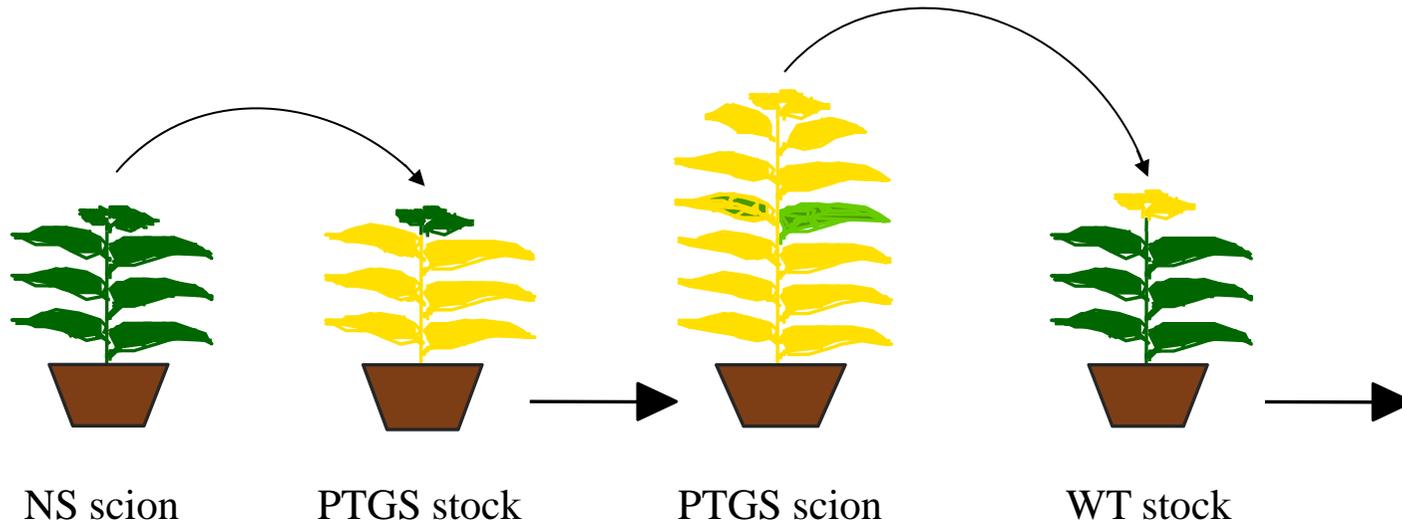
PTGS produces a sequence-specific systemic silencing signal



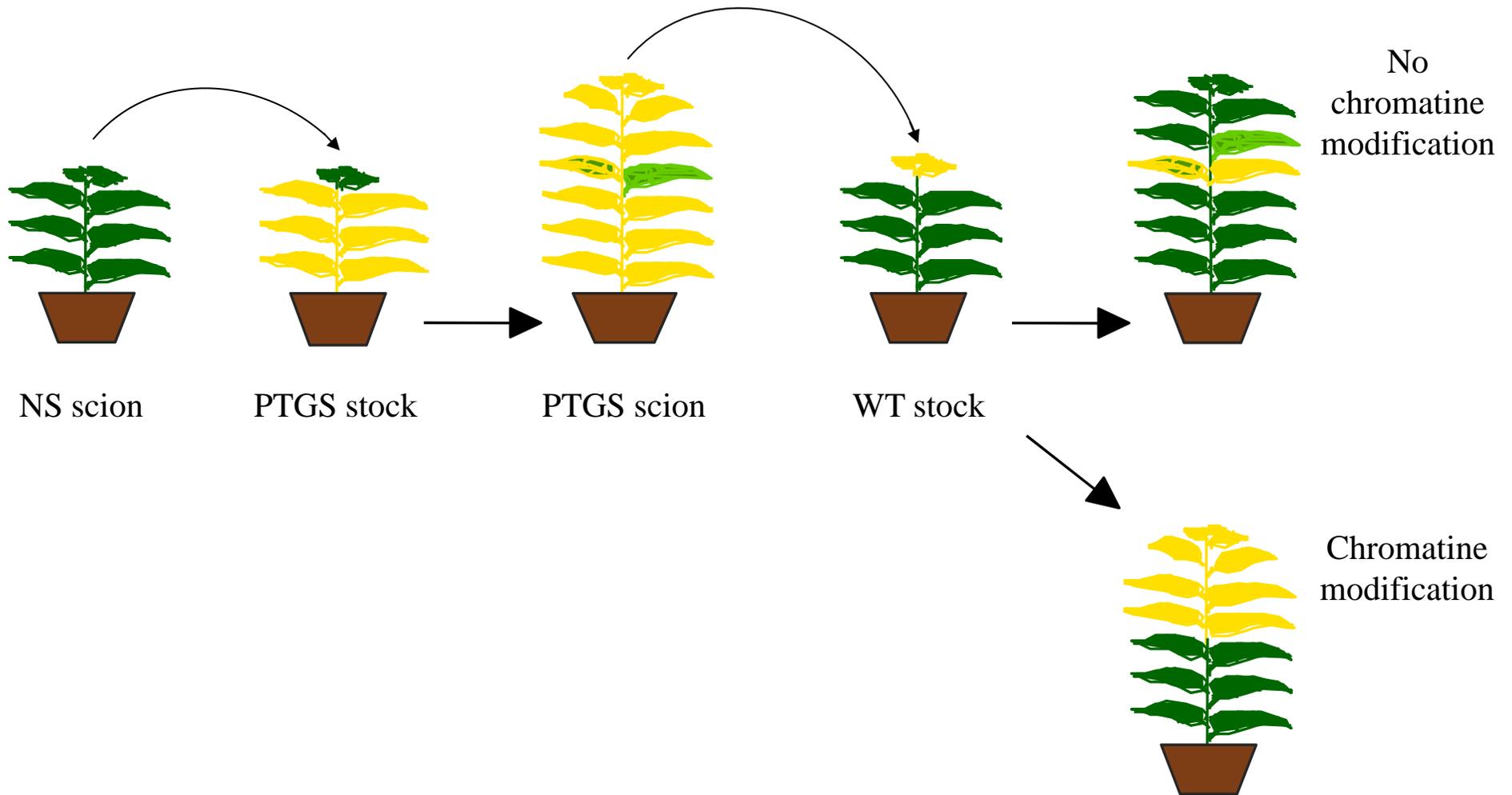
PTGS produces a sequence-specific systemic silencing signal



PTGS maintenance requires chromatin modification

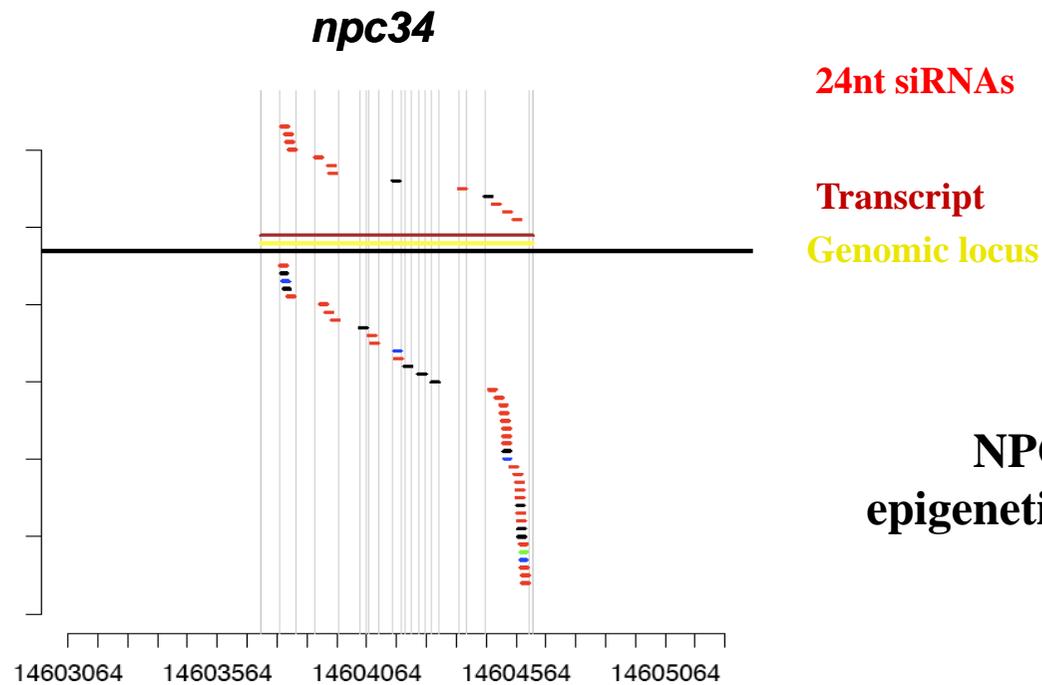


PTGS maintainance requires chromatine modification



RNA precursors of DCL3-dependent 24-nt siRNAs are labile

Very few have been cloned

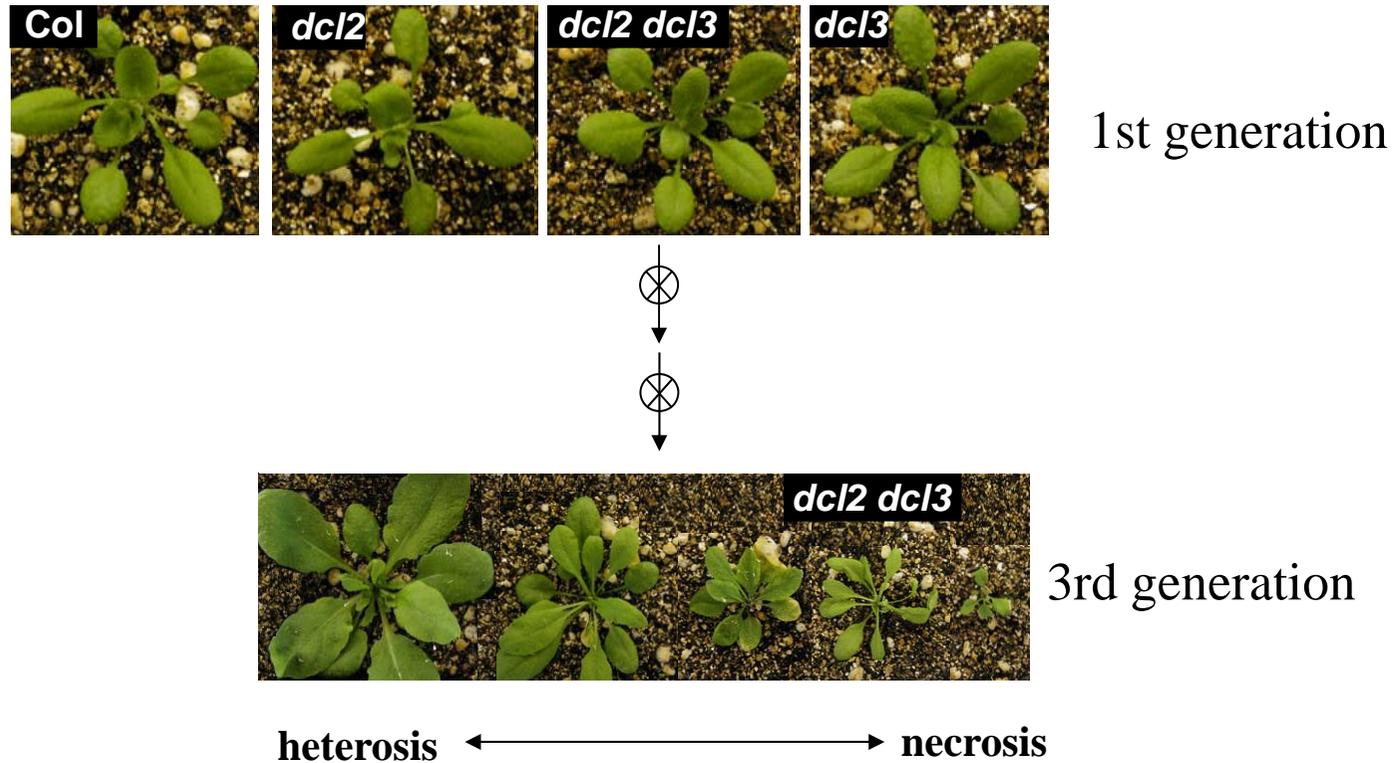


NPC34 chromatin is marked by epigenetic marks linked to PcG repression

DCL2 and DCL3 act redundantly to prevent genomic shocks

dcl2 (Col) x *dcl3* (Col) -> viable *dcl2 dcl3*

dcl2 (Ws) x *dcl3* (Col) -> epigenetically unstable *dcl2 dcl3*



LabEx project 3 : Epigenetics and RNA-mediated regulation

Is it possible to specifically modify the RNA repertoire by modifying the RNA machinery and/or its cellular localization to improve plants' epigenetic potential ?

Work packages

In this project we will explore epigenetic mechanisms using a combination of cell biology , genetics and genomics.

1. Molecular identification of epigenetic targets

2. Subcellular architecture and trafficking dynamics of non-coding RNAs

People involved :

ISV Lab: RNA regulators

M. Crespi

C. Hartmann

C. Sorin

C. Lelandais

A. Christ

F. Ariel

F. Bardou

A. Moreno

P. Bustos

IJPB Lab: Epigenetics and Small RNA

H. Vaucheret

T. Elmayan

I. Le Masson

C. Albertini

N. Bouteiller

E. Elvira-Matelot

E. Martinez de Alba

J-S Parent

N. Shamandi

Bioinformatics : Daniel Gautheret at Orsay-PXI and
Hadi Quesneville / Matthias Zynnicky at URGI